

Grant Information: Institution, Principal Investigator(s), Contact Information, Grant Number	University of Massachusetts Project: A Novel Strategy for Arsenic Phytoremediation Project Leaders: <u>Om Parkash Dhankher</u> , <u>Venkataraman Dhandapani</u> , <u>Jason C. White</u> , <u>Baoshan Xing</u> Grant Number: R01ES032686 Funding Period: 2021-2025
Technology	Developing a genetics-based phytoremediation strategy for arsenic uptake, translocation, detoxification, and hyperaccumulation into the fast-growing, high biomass, non-food oilseed crop <i>Crambe abyssinica</i> .
Innovation	<b>Materials:</b> Nanosulfur will be utilized to modulate the bioavailability and phytoextraction of arsenic from soil and to increase the storage capacity via enhanced sulfur assimilation.
	crop <i>Crambe abyssinica</i> to remediate arsenic-contaminated soil.
	Why is this technology/approach different than what is already in the market? We are using a gene pyramiding approach to co-express several genes that control the transport, oxidation state, and binding of As for efficient extraction and hyperaccumulation into above-ground plant tissues of <i>Crambe abyssinica</i> . Phytoremediation is a cost-effective and ecologically friendly alternative to physical remediation methods.
Contaminant and Media	<b>Contaminants:</b> Arsenic <b>Media:</b> Soil, sediment, and maybe water
Expansion Potential	Looking Forward: What other contaminants/media would work for your technology? Toxic metals: Pb, Cd, Hg, Cr Combined Remedy: Would this technology work well with other treatment approaches? Yes, this approach could be combined with biofuels production on contaminated sites.
Sites/Samples	We are using artificially contaminated soils, but will use the real-world samples, like field soils contaminated with As and other toxic metals.

Continued



Technology Readiness Level	TRL 3 — Experimental proof of concept TRL 4 — Technology validated in laboratory
Update of Progress	<ul> <li>We are co-expressing four genes that control the transport, oxidation state, and binding of As for efficient extraction and hyperaccumulation into above-ground plant tissues. All four genes are cloned and transformed into <i>Crambe</i> either single or stacked genes.</li> <li>We have already developed transgenic <i>Crambe</i> plants for several gene constructs.</li> <li>Analysis of double transgenic lines coexpressing arsenate reductase and glutathione biosynthesis pathway gene showed that double transgenic plant had significantly increased arsenate (AsV) tolerance as these plants attained</li> </ul>
	almost three-fold higher biomass compared to wild type controls plants. These plants accumulated 2-fold more arsenic in the shoot tissues. Analysis of plants expressing other genes is in progress.
	<ul> <li>Analysis of transgenic plants for other genes is in progress.</li> </ul>
	<ul> <li>We have also optimized the nanosulfur concentration on arsenic mobility from soil and subsequent uptake and accumulation in <i>Crambe</i> grown in soil supplemented with both arsenate and nanosulfur.</li> </ul>
	<ul> <li>Additionally, for the Diversity Supplement award, we are modulating the expression of arsenate reductases for increasing tolerance and reducing As accumulation in rice for food safety.</li> </ul>



Developing a genetics-based strategy for arsenic phytoremediation in Crambe abyssinica, a non-food industrial oilseed crop.